

# The Environmental Effects of Sea Pollution on Microorganisms

Musa Kalsom, Binazeer Mehman

Department of Biology, Faculty of Science, University of Pishavear, Pishavear, Pakistan

Received: 21 September 2020

Accepted: 03 November 2020

Published: 01 December 2020

## Abstract

Sea anemones from phylum cnidaria are benthic organisms and live on sea substrates, so water pollution can affect them easily. The marine environments are the target of contaminations caused by a complex mixture of metals and chemical materials from various anthropogenic sources. The pollution could have mortal effects on sea anemones. Although the anemones respond to increased metal exposure by closing their tentacles and changing the activities of the enzymes, CAT (catalase), GR (glutathione reductase), and CA (carbonic anhydrase), altered enzyme activity and tentacle retraction of sea anemones, as well as decreased zooxanthellae cell density could be observed responses over exposure. Metal depuration and physiological recovery are dependent on both the metal and the exposure concentration. Copper exposure can cause tentacle retraction and increase mucus production in both symbiotic and aposymbiotic anemones. Heavy metals especially copper can cause cellular damages in the level of nucleus and DNA. Some species of sea anemones accumulate heavy metals such as copper and are sensitive, as effects were detected at environmentally relevant copper concentrations. Likewise, they may be useful in biomonitoring copper polluted environments.

**Keywords:** Sea anemones; Contaminants; Antioxidant Enzymes; DNA Damage; Bioaccumulation; Symbiosis Relationship

## How to cite the article:

M. Kalsom, B. Mehman, *The Environmental Effects of Sea Pollution on Microorganisms*, *Medbiotech J.* 2020; 4(4): 161-168, DOI: 10.22034/mbt.2020.122172

©2020 The Authors. This is an open access article under the CC BY license

## 1. Introduction

Sea anemones are polyp like and solitary organisms which absorb dissolved organic matters as food or have symbiotic life with photosynthetic zooxanthellates. Being not much mobile make them feed on planktons [1]. Knowing about the reaction of these organisms against pollutions is important due to their little mobility and disability in escaping [2]. Despite their importance in the coastal environment worldwide, there are few studies related to metal accumulation in the tissues of cnidarians and even less in sea anemones [3,4]. Marine environment are contaminated via many different forms, such as solid waste (e.g., plastics), increased nutrient (e.g., nitrates and phosphates),

toxic chemicals (e.g., organic compounds, DDT, PCB, metals, pharmaceuticals, gas), oil spills and sediment inputs due to human activities (e.g., industry, agriculture, on deforestation, sewage discharge, aquaculture), radioactivity, and discarded fishing nets. These contaminations change the biological, chemical and physical and characteristics of the oceans and coastal zones, and potentially threatens marine organism, ecosystems, and biodiversity and affects thus the quality and productivity of marine ecosystems. The final effect of pollution on marine resources and different organisms depend on the intensity (acute or chronic), form of pollution, and location of the contamination. Also there is an important point

that some marine environments, ecosystems, and species are more sensitive than others to pollution [5].

As noted above, heavy metals are important source of contamination. The term "heavy metals" is defined for any metallic element that has a relatively high density and is toxic or poisonous even at low concentration [6]. Heavy metals include lead (Pb), cadmium (Cd), zinc (Zn), mercury (Hg), arsenic (As), silver (Ag) chromium (Cr), copper (Cu) iron (Fe), and the platinum group elements [7]. Due to input of metal pollution from different ways, marine environments contain a complex mixture of metals. However, few studies have noted the effects of metal mixtures on sea anemones. Despite of this fact, anemones have been shown to be sensitive to metal exposure [4, 8-9].

Global climate changes, such as ocean acidification (OA) have intensified the effects of these local stressors. Anthropogenic activities like fossil fuel combustion, cement production and deforestation have significantly increased atmospheric Carbon dioxide (CO<sub>2</sub>) levels [10, 11]. The ocean pH decreases by increasing of CO<sub>2</sub> and it rises the hydrogen ion concentration in water [12].

Presence and absence of hydroxyl (OH<sup>-</sup>) and carbonate ions (CO<sub>3</sub><sup>2-</sup>) change the fate of metals in the aquatic environments [13]. When the effects of both stressors (CO<sub>2</sub> and metals) combine, as well as the increased bioavailability of metals at lower pH levels, substantial biological effects might be at the physiological level, which may result in having repercussions for populations and even ecosystems [14-16].

Cu is a commonly used metal that can enter marine ecosystems through different ways such as sewage treatment discharge, industrial effluent, leaching of anti-fouling paints, and copper refineries [3, 17, 18], metal mining [19]. Marine cnidarians would be under oxidative stress by relatively low levels of Cu [20-22].

The sea anemone, *Aiptasia pallida*, is a symbiotic cnidarian, which is endemic to the southeastern United States and its habitat is commonly on hard substrates such as coral reefs and rocks [23][24]. The photosynthetic zooxanthellae symbiont earns protection, nitrogen, and a carbon dioxide source from *A. pallida* in exchange for energy [25]. Environmental stressors such as temperature change and contaminants may interrupt this relationship, potentially causing loss of the zooxanthellae, resulting in bleaching of *A. pallida* [26-27].

Some physiological effects have been well studied on marine cnidarians as a consequence of Cu exposure [4][22][28][29]. In particular, Cu has been shown to cause the generation of reactive oxygen species (ROS), which can lead to degradation of macro-molecules like proteins,

lipids and DNA [30][31]. Several anti-oxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione reductase (GR), and glutathione peroxidase (GPx) [31] are used to combat the harmful effects of ROS. The activity of these enzymes as well as carbonic anhydrase (CA), responsible for acid/base balance, were altered in cnidarians exposed to Cu in previous studies [4, 20, 21, 32].

When an imbalance between the number of reactive oxygen species formed and the neutralization of these by antioxidant defenses is occurred, some type of damage and changes in cellular signals, it is said that the cell is under oxidative stress [33,35,36]. There is evidence in marine organisms that the toxicity of copper causes oxidative stress, an increase in antioxidants and DNA damage [37].

Carbonic anhydrase (CA) is an essential enzyme, catalyzes the interconversion of carbon dioxide to bicarbonate and is important in respiration and metabolism [38]. The metalloprotein and enzyme CA pivots around zinc as its catalytic redox center and is known to be sensitive to metal exposure. Metals have been shown to inhibit normal CA activity in fish, blue crabs, and recently corals and sea anemones [20, 39-41].

The sea anemone *A. pallida* are abundant in shallow temperate and tropic coastal environments. Also their sampling and compatibility to laboratory conditions is easy. These organisms reproduce quickly compared to other symbiotic cnidarians and their habitat is mostly near contaminated marine zones, so are good model organisms for toxicological studies [42]. This species has also been shown to be sensitive to several different metals, particularly Cu [4, 8, 9].

The induction of multiple xenobiotics can rises populational or individual resistance in contaminated waters. Therefore, evaluation of the level of xenobiotics induction in individuals could be used as a biomarker of capability in these organisms [43]. Of course, for better perception of the biological responses to stressors, both genetic and cellular responses should be integrated, and physiological responses should be analyzed from an ecotoxicological perspective [44, 45].

An inhibition in the extrusion mechanism of contaminants might result in roxidative stress, and can then generate genetic damage, which is often irreversible. In the relevant literature, one may find research involving copper, oxidative stress and DNA damage [46]; copper and xenobiotics in coral [47], lead and xenobiotics in bivalves [48]; however, finding researches using metals in invertebrates, showing oxidative stress, and multiple xenobiotic expression linked to DNA damage are not common.

Sea anemones are benthic marine organisms with less mobility that escape or immigration could not be a good method of dealing with pollution. Therefore physiological, biological and genetic responses and changes would be signs of alternations by different contaminants. The structure and the cellular mechanisms of *Bunodosoma caissarum* enables it to support extreme conditions, such as air exposure and fluctuations in salinity. This species has different mechanisms of adaptation for coping with stress situations, including mucus secretion, the presence of warts and the development of a protective dome shape [49].

## 2. Marine pollution on enzymatic changes:

Changes in the activity of antioxidant enzymes is one of the physiological effects of metal exposure. Carbonic anhydrase (CA) activity could be an important biomonitoring tool for assessing cnidarian health. Although some research has been conducted to elucidate the effects of individual metal exposure to sea anemones [4, 8, 9], little is known about the impact of metal mixtures.

When sufficient light is provided photosynthetic zooxanthellae can generate reactive oxygen species (ROS), which can cause cellular and molecular damage, such as oxidation of phospholipid membranes, denaturation of proteins, and alteration of nucleic acids [50-52]. The cell membrane, is one of the centers of reactive oxygen species (ROS) activity. In biological systems, Cellular damage results basically from ROS attack on macromolecules such as sugars, DNA, proteins and lipids [34, 53, 54]. One way cells can avoid ROS generation and the resulting oxidative stress would be through the extrusion of the contaminants.

When abiotic stress increases, electron transport mechanisms (i.e., photosystems and mitochondrial transport chains) become less efficient. In turn, free electrons are more likely to interact with diatomic molecular oxygen ultimately resulting in a series of reduced reactive oxygen species (ROS). Both *Exaiptaisa pallida* and its symbiotic zooxanthellae have evolved mechanisms to protect against the damaging effects of ROS [51][52]. Due to photosynthesis, symbiotic anemones are subjected to very high levels of oxygen during the day, and at night they experience a hypoxic internal environment [55]. This continuous transition between extreme oxygen environments can result in the increased production of ROS [55][56]. ROS can denature proteins, mutate DNA, and cause lipid peroxidation [51]. To neutralize the potentially harmful effects of ROS, organisms produce enzymes such as superoxide dismutase (SOD) and catalase (CAT).

SOD catalyzes the conversion of superoxide anion into hydrogen peroxide and oxygen; whereas CAT

catalyzes the conversion of hydrogen peroxide ( $H_2O_2$ ) into water and oxygen [57].  $H_2O_2$  can be induced by a variety of factors including metal exposure [58]. In the presence of metal ions such as Cu,  $H_2O_2$  can be quickly converted to hydroxyl radicals in a process known as the Fenton reaction [59]. Zn is one of other heavy metals can also cause oxidative stress and hydroxyl radical formation via different mechanisms.

Through another enzymatic pathway, glutathione peroxidase (GPX) can also catalyze the conversion of  $H_2O_2$  to water, using monomeric glutathione as the reducing agent [60-63]. A subsequent reaction reduces the glutathione using NADPH and glutathione reductase (GR) as the catalyst, so that it may be recycled for the previous reaction [60-63]. Glutathione itself can also function in metal detoxification by binding to and removing metals from the organism via conjugation reactions [60]. Cellular destruction on the level of DNA damages by pollutants:

Copper, even at low concentrations and even within the limit allowed by Brazilian law, causes stress in anemone cells. This metal exhibits cytotoxicity, leading cells to activate cellular defense mechanisms to some extent (in lower concentration). This defense is not sufficiently effective to prevent an increase of reactive oxygen species and genetic damage, however, when it is not activated, (in the highest concentration) the cytotoxic and oxidative genetic damage, are even greater [2].

On the work of Schwarz and Collaborators (2013) it was seen that there is an increase in DNA damage in cells exposed to copper, with a clear increase in the percentage of genetic material in the tail. Copper has the potential to cause DNA damage, especially in relation to dispersion of the genetic material (larger DNA% than tail size, itself), which seems to show simple breaks in DNA chains [64]. It is noteworthy that the percentage of DNA present in the tail seems to be, among the parameters analyzed at Comet assay, the most sensitive, as it was the analysis that managed to show damage in 6 h of exposure, even in the copper concentration 7.8 lg/L. The occurrence of xenobiotic contaminants, such as metal ions, in the marine environment is highly harmful to the biological integrity as well as the function of marine organisms. Many of these pollutants are chemicals capable of causing damage to DNA, usually through oxidative stress. Benzo[a]pyrene, for example, increases the amount of ROS which leads to formation, directly or indirectly, of DNA adducts, resulting in genetic chain breaks [65-66]. Bopp and Collaborators (2008), in addition to investigating the formation of reactive oxygen species in gill cells exposed to copper, also performed comet assays, to analyze

DNA integrity in cells [67]. Generation of ROS's are essential factors for the DNA damage, with the reactive oxygen species performing a crucial role in the process of DNA degradation, causing direct damage through oxidation of DNA molecule, or indirect, causing damage to lipids and proteins, subsequently leading to genotoxic damage [46,67-71].

Color and symbiotic relationship disorders by contaminants:

Due to close vicinity of *Aiptasia pallida* to anthropogenic inputs, may be a bioindicator of copper pollution. Biomonitoring organisms can indicate the quality of the environment they inhabit. These organisms can be used to understand the temporal and geographical variations in the bioavailability of contaminants [72]. The color of a healthy *A. pallida* is golden-brown in color due to their zooxanthellae (dinoflagellate algae of the genus *Symbiodinium*), which live in symbiosis with *A. pallida* [73]. In recent years, pulse-amplitude modulated (PAM) fluorometry has been used to assess the effects of pollutants on seaweed, algae, and zooxanthellae [74-76]. The PAM fluorometer can be used to distinguish between photon energy captured by a chlorophyll-a pigment molecule used to drive photosynthesis, and energy emitted as fluorescence or converted to heat [77]. In this symbiotic relationship *A. pallida*, provides the zooxanthellae with nutrients (nitrogenous waste and CO<sub>2</sub>) and protection. In return, zooxanthellae provide the anemone with photosynthate via carbon fixation pathways [77]. When this host-symbiont relationship becomes challenged by any number of environmental stressors, loss of the symbiont "bleaching" and color loosing may occur [78]. Bleaching of sea anemones may reduce tissue metal concentrations, and has been demonstrated in studies as a consequence of copper exposure [4, 21].

Corals and anemones both have been studied after copper exposure and tissue copper accumulation [8, 9, 79]. The pattern of copper accumulation in the tissues of symbiotic cnidarians may be affected by the presence of the symbiont. Copper accumulation in the symbiotic sea anemone, *Anemonia viridis*, was not directly reflective of the copper concentrations in the environment, suggesting that detoxification and excretion mechanisms were used to combat increasing copper concentrations [79]. Harland and Nganro (1990) also suggested that greater extent copper is accumulated in the zooxanthellae comparison to host tissue and in a final detoxification attempt the anemone release their symbiotic algae to reduce internal copper concentrations [80]. The concept of zooxanthellae expulsion as a mechanism of metal detoxification has been suggested by several researchers, since zooxanthellae have been found to accumulate

heavy metals to a larger extent and be more tolerant than their symbiotic hosts [81,82].

Bioaccumulation as a response to pollution:

The sea anemones *Bunodosoma cangicum* and *Bunodosoma caissarum* are commonly Brazilian endemic species found in the intertidal area of the Southeast regions of Brazil [49, 83]. Organisms from this species can bioaccumulate heavy metals from the environment [84]. *Bunodosoma caissarum* is capable of combining Hg and other metals such as cadmium, copper and zinc in its tissues [85].

In two microcosms with and without sea anemone *B. caissarum* different results were achieved in the study of Rizzini Ansari et al. (2015). In both microcosms, methylmercury (MeHg) formation was observed. A higher level of MeHg was detected in the seawater from the microcosm without *B. caissarum*. The reason for this might be that under these conditions the microorganisms were allowed to grow and to contribute to MeHg production without the Hg bioaccumulation interference of *B. caissarum*. Another possibility is that *B. caissarum* readily incorporated part of the MeHg produced. Mercury was probably less available for methylation in microcosms with *B. caissarum* because of bioaccumulation and due to the higher concentrations of suspended particulate matter that could form complexes with Hg. This experiment demonstrated the formation of MeHg in the seawater of microcosms without *B. caissarum*. However, we should note that in the microcosms containing *B. caissarum* this experiment does not permit a conclusion as to whether MeHg was formed in the specimens themselves, in the seawater, or in the mucus secretions and then accumulated by *B. caissarum* [85].

### 3. Conclusion

The copper represents an adversity to the anemones cells, being cytotoxic and genotoxic. Metal accumulation in the tissues of sea anemones are a good indicator of metal exposure at different testable concentrations. Understanding how sea anemones and their symbionts are affected by metal exposures in the laboratory may allow better understanding about the responses of symbiotic cnidarians in metal polluted aquatic environments. The Symbiont loss, as might be observed by decreased algal cell density and visible paling, might be aided in reducing the tissue metal burden of the anemones.

Additionally metal accumulation patterns in the anemones, marked differences were observed in responses of antioxidant enzymes to a mixed metal exposure regime as compared to individual metal exposure. The activity of CAT, for example, increased after exposure to Cu alone [4], and decreased as a consequence of mixed metal

exposure, which may also be a consequence of the initial reduction in zooxanthellae cell density

These organisms could be susceptible to metal pollution in the environment, particularly along the coastline surrounding heavily populated areas with substantial anthropogenic inputs. Symbiotic sea anemone upregulates CAT activity to combat the damaging effects of hydrogen peroxide. SOD activity significantly decreases during the highest copper exposure. SOD is substantially reduced in aposymbiotic *A. pallida*, suggesting that the zooxanthellae are associated with the oxidative stress response. *A. pallida* may be a useful bioindicator species in laboratory studies and potentially in metal polluted environments. Sea anemones may face to Reduced zooxanthellae electron transport, Reduced carbonic anhydrase activity and Reduced growth when they exposure to contaminants.

## References

- 1) de Capitani, J.D. 2007. Estrutura populacional e variabilidade genética de anêmonas-do-mar da região entremares de costão rochoso," Dissertação (mestrado) – Universidade Estadual de Campinas, Instituto de Biologia.
- 2) Anjos, V. A., da Silva-Junior, F. M. R., and Souza, M. M. 2014. Cell damage induced by copper: An explant model to study anemone Cells. *Toxicology in Vitro*, 28: 365–372.
- 3) Mitchelmore, C.L., Alan Verde, E., Ringwood, A.H., and Weis, V.M. 2003. Differential accumulation of heavy metals in the sea anemone *Anthopleura elegantissima* as a function of symbiotic state, *Aquatic Toxicology*, 64: 317– 329.
- 4) Main, W. P. L., C. Ross, C., and Bielmyer, G. K. 2010. Copper accumulation and oxidative stress in the sea anemone, *Aiptasia pallida*, after waterborne copper exposure. *Comparative Biochemistry and Physiology*, 151: 216–221.
- 5) Wilhelmsson, D., Thompson, R. C., Holmstrom, K., Linden, O., and Eriksson-Hagg, H. 2013. Managing ocean environment in a changing climate, chapter 6, *Marine Pollution*, pp. 127-169.
- 6) Lenntech Water Treatment and Air Purification (2004). Water Treatment, Published by Lenntech, Rotterdamseweg, Netherlands, (www.excelwater.com/thp/filters/Water-Purification.htm).
- 7) Duruibe, J. O., Ogwuegbu, M. O. C., and Egwurugwu, J. N. 2007. Heavy metal pollution and human biotoxic effects, *International Journal of Physical Sciences*, 2: 112-118.
- 8) Mitchelmore, C. L., Ringwood, A. H., Weis, V. M. 2003. Differential accumulation of cadmium and changes in glutathione levels as a function of symbiotic state in the sea anemone *Anthopleura elegantissima*, *Journal of Experimental Marine Biology and Ecology*, 284: 71 – 85.
- 9) Mitchelmore, C. L., Verde, E. A., Weis, V. M. 2007. Uptake and partitioning of copper and cadmium in the coral *Pocillopora damicornis*, *Aquatic Toxicology*, 85: 48–56.
- 10) Canadell, J. G., Le Quere, C., Raupach, M. R., Field, C. B., Buitenhuis, E. T., Ciais, P., Conway, T. J., Gillett, N. P., Houhton, R. A., Marland, G. 2007. Contribution to accelerating atmospheric CO<sub>2</sub> growth from economic activity, carbon intensity, and efficiency of natural sinks. *Proceeding of the National Academy of Sciences of the United States of America*, 104: 18866–18870.
- 11) Yool, A., Popova, E. E., Coward, A. C., Bernie, D., Anderson, T. R. 2013. Climate change and ocean acidification impacts on lower trophic levels and the export of organic carbon to the deep ocean, *Biogeosciences*, 9: 5831– 5854.
- 12) Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. A., Gnanadesikan, A., Gruber, N., Ishida, A., Joos, F., Key, R. M., Lindsay, K., Maier-Reimer, E., Matear, R., Monfray, P., Mouchet, A., Najjar, R. G., Plattner, G. K., Rodgers, K. B., Sabine, C. L., Sarmiento, J. L., Schlitzer, R., Slater, R. D., Totterdell, I. J., Weirig, M. F., Yamanaka, Y., Yool, A. 2005. Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms, *Nature*, 437: 681–686.
- 13) Millero, F. N. J., Woosely, R., DiTrollo, B., Waters, J. 2009. Effects of ocean acidification on the speciation of metals in seawater. *Oceanography* 22: 72–85.
- 14) Langdon, C., Takahashi, T., Sweeney, C., Chipman, D., Goddard, J., Marubini, F., Aceves, H., Barnett, H., Atkinson, M. J. 2000. Effect of calcium carbonate saturation state on the calcification rate of an experimental coral reef, *Global Biogeochemical Cycle*, 2: 639–654.
- 15) Leclercq, N., Guttuso, J. P., Jaubert, J. CO<sub>2</sub> partial pressure controls the calcification rate of coral community, 6: 329–334.
- 16) Le Quesne, W. J. F., and Pinnegar, J. K. 2012. The potential impacts of ocean acidification: scaling from physiology to fisheries, *Fish and Fisheries*. 13: 333–334.
- 17) Guzmán, H. M., and Jiménez, C. E. 1992. Contamination of coral reefs by heavy metals along the Caribbean coast of Central America, *Marine Pollution Bulletin*, 11: 554–561.
- 18) Jones, R. J. 1997. Zooxanthellae loss as a bioassay for assessing stress in corals, *Marine Ecology Progress Series Journal*, 149: 163–171.
- 19) Bryan, G. W., Some aspects of heavy metal tolerance in aquatic organisms, In: Lockwood, A.P.M. (Ed.). 1974. *Effects of Pollutants on Aquatic Organisms*. University Press, Cambridge, England, pp. 7–34.
- 20) Bielmyer, G. K., Grosell, M., Bhagooli, R., Baker, A. C., Langdon, C., Gillette, P., Capo, T. R. 2010. Differential effects of copper on three species of

- scleractinian corals and their algal symbionts (Symbiodinium spp.), *Aquatic Toxicology* 97: 125–133, 2010.
- 21) Brock, J. R., and Bielmyer, G. K. 2013. Metal accumulation and sublethal responses in the sea anemone, *Aiptasia pallida* after waterborne exposure to metal mixtures. *Comparative Biochemistry and Physiology Part C*, 158: 150–158
- 22) Patel, P. P., and Bielmyer-Fraser, G. K. 2015. The influence of salinity and copper exposure on copper accumulation and physiological impairment in the sea anemone, *Aiptasia pallida*, *Comparative Biochemistry and Physiology Part C*, 168: 39–47.
- 23) Kaplan, E. H. 1988. A Field Guide to Southeastern and Caribbean Seashores, Peterson Field Guides. Houghton Mifflin Company, Boston, MA 213.
- 24) Shick, J. M., 1991. A Functional Biology of Sea Anemones, Chapman & Hall, London, UK.
- 25) Goulet, T. L., Cook, C. B., and Goulet, D. 2005. Effect of short-term exposure to elevated temperatures and light levels on photosynthesis of different host-symbiont combinations in the *Aiptasia pallida* Symbiodinium Symbiosis, *Limnology Oceanography*, 50: 1490–1498.
- 26) Mercier, A., Pelletier, É., and Hamel, J. 1997. Effects of butyltins on the symbiotic sea anemone *Aiptasia pallida* (Verrill), *Journal of Experimental Marine Biology and Ecology*, 215: 289–304.
- 27) Perez, S. F., Cook, C. B., Brooks, W. R. 2001. The role of symbiotic dinoflagellates in the temperature-induced bleaching response of the subtropical sea anemone *Aiptasia pallida*, *Journal of Experimental Marine Biology and Ecology*, 256: 1–14.
- 28) Howe, P., Reichelt-Brushett, A. J., and Clark, M. 2012. *Aiptasia pulchella*: a tropical cnidarian representative for laboratory ecotoxicological research. *Environmental Toxicology and Chemistry*, 11: 2653–2662.
- 29) Howe, P., Reichelt-Brushett, A. J., Clark, M. W. 2014. Effects of Cd, Co, Cu, Ni, and Zn on the asexual reproduction and early development of the tropical sea anemone *Aiptasia pulchella*, *Ecotoxicology*, 23: 1593–1606.
- 30) Chang, L. W., Magos, L., Suzuki, T. (Eds.), Lewis/CRC Publishers, Boca Raton, pp. 1–1198. 1996.
- 31) Luschnik, V.I. 2011. Environmentally induced oxidative stress in aquatic animals. *Aquatic Toxicology*, 101: 13–30.
- 32) Siddiqui, S., Goddard, R. H., and Bielmyer-Fraser, G. K. 2015. Comparative effects of dissolved copper and copper oxide nanoparticle exposure to the sea anemone, *Aiptasia pallida*, *Aquatic Toxicology*, 160: 205–213, 2015.
- 33) Bandyopadhyay, U., Das, D., and Banerjee, R. K. 1999. Reactive oxygen species: oxidative damage and pathogenesis, *Current Science*, 77: 658–666, 1999.
- 34) Berra, C. M., Menck, C. F. M., and Di Mascio, P. 2006. Estresse oxidativo, lesões no genoma e processos de sinalização no controle do ciclo celular, *Quim. Nova*. 29: 1340–1344.
- 35) Lesser, M. 2012. Oxidative stress in aquatic ecosystems. In: *Oxidative Stress in Tropical Marine Ecosystems*, Blackwell Publishing Ltd., 9–19.
- 36) Paital, B., and Chainy, G. B. N. 2012. Effects of salinity on O<sub>2</sub> consumption, ROS generation and oxidative stress status of gill mitochondria of the mud crab *Scylla serrata*, *Comparative Biochemistry and Physiology Part C*, 155: 228–237.
- 37) Lee, J. A., Marsden, I. D., and Glover, C. N. 2010. The influence of salinity on copper accumulation and its toxic effects in estuarine animals with differing osmoregulatory strategies, *Aquatic Toxicology*, 99: 65–72.
- 38) Gilmour, K. M. 2010. Perspectives on carbonic anhydrase. *Comparative Biochemistry and Physiology Part A*, 157: 193–197.
- 39) Gilbert, A. L., and Guzmán, H. M. 2001. Bioindication potential of carbonic anhydrase activity in anemones and corals, *Marine Pollution Bulletin*, 42: 742–744.
- 40) Skaggs, H. S., and Henry, R. P. 2002. Inhibition of carbonic anhydrase in the gills of two euryhaline crabs, *Callinectes sapidus* and *Carcinus maenas*, by heavy metals, *Comparative Biochemistry and Physiology Part C*, 133: 605–612.
- 41) Morgan, T. P., Grosell, M., Gilmour, M., Playle, R. C., and Wood, C. M. 2004. Time course analysis of the mechanism by which silver inhibits Na<sup>+</sup> and Cl<sup>-</sup> uptake in gills of rainbow trout, *American Journal of Physiology Regulatory Integrative Comparative Physiology*, 287: 234–242.
- 42) Leal, M. C., Nunes, C., Engrola, S., Dinis, M. T., and Calado, R. 2012. Optimization of monoclonal production of the glass anemone *Aiptasia pallida* (Agassiz in Verrill, 1864), *Aquaculture*, 91–96.
- 43) Kurelec, B., Smital, T., Pivcevic, B., Eufemia, N., and Epel, D. 2000. Multixenobiotic resistance, P-Glycoprotein and chemosensitizers, *Ecotoxicology*, 9: 307–327.
- 44) Spurgeon, D. J., Ricketts, H., Svendsen, C., Morgan, A. J., and Kille, P. 2005. Hierarchical responses of soil invertebrates (earthworms) to toxic metal stress, *Environmental Science and Technology*, 39: 5327–5334.
- 45) Regoli, F., Gorbi, S., Frenzilli, G., Nigro, M., Corsi, I., Focardi, S., and Winston, G. W. 2002. Oxidative stress in ecotoxicology: from the analysis of individual antioxidants to a more integrated approach, *Marine Environmental Research*, 54: 419–423.
- 46) Schwarz, J. A., Mitchelmore, C. L., Jones, R., O'Dea, A., and Seymour, S. 2013. Exposure to copper induces oxidative and stress responses and

- DNA damage in the coral *Montastraea franksi*, *Comparative Biochemistry and Physiology Part C*, 157: 272–279.
- 47) Venn, A. A., Quinn, J., Jones, R., and Bodnar, A. 2009. P-glycoprotein (multi-xenobiotic resistance) and heat shock protein gene expression in the reef coral *Montastraea franksi* in response to environmental toxicants, *Aquatic Toxicology*, 93: 188–195.
- 48) Rocha, C. T., and Souza, M. M. 2012. The influence of lead on different proteins in gill cells from the freshwater bivalve, *Corbicula fluminea*, from defense to repair biomarkers, *Archive of Environmental Contamination and Toxicology*, 62: 56–67.
- 49) Amado, E. M., Vidolin, D., Freire, C. A., Souza, M. M. 2011. Distinct patterns of water and osmolyte control between intertidal (*Bunodosoma caissarum*) and subtidal (*Anemonia sargassensis*) sea anemones, *Comparative Biochemistry and Physiology Part A*, 158: 542–551.
- 50) Jamieson, D., Chance, B., Cadenas, E., and Boveris, A. 1986. The relationship of free radical production to hyperoxia. *Annual Review of Physiology*, 48: 703–719.
- 51) Richier, S., Furla, P., Plantivaux, A., Merle, P., and Allemand, D. 2005. Symbiosis-induced adaptation to oxidative stress, *Journal of Experimental Biology*, 208: 277–285.
- 52) Lesser, M. P. 2006. Oxidative stress in marine environment: biochemistry and physiological ecology, *Annual Review of Physiology*, 68: 253–278.
- 53) Sandrini, J. Z., Bianchini, A., Trindade, G. M., Nery, L. E. M., and Marins, L. F. F. 2009. Reactive oxygen species generation and expression of DNA repair-related genes after copper exposure in zebrafish (*Danio rerio*) ZFL cells, *Aquatic Toxicology*, 95: 285–291.
- 54) Vasconcelos, S. M. L., Goulart, M. O. F., Moura, J. B. F., Manfredini, V., and Benfato, M. S. 2007. Espécies reativas de oxigênio e de nitrogênio, antioxidantes e marcadores de dano oxidativo em sangue humano: principais métodos analíticos para sua determinação, *Quim. Nova*, 30: 1323–1338.
- 55) Dykens, J. A., Shick, J. M., Benoit, C., Buettner, G. R., and Winston, G. W. 1992. Oxygen radical production in the sea anemone *Anthopleura elegantissima* and its endosymbiotic algae, *Journal of Experimental Biology*, 168: 219–241.
- 56) Nii, C. M., and Muscatine, L. 1997. Oxidative stress in the symbiotic anemone *Aiptasia pulchella* (Carlgren, 1943): contribution of the animal to superoxide ion production at elevated temperatures, *Biology Bulletin*, 192: 444–456.
- 57) Higuchi, T., Fujimura, H., Hitomi, Y., Arakaki, T., Oomori, T., and Suzuki, Y. 2010. Photochemical formation of hydroxyl radicals in tissue extracts of the coral *Galaxea fascicularis*, *Photochemical and Photobiological Sciences*, 86: 1421–1426.
- 58) Cabisco, E., Piulats, E., Echave, P., Herrero, E., and Ros, J. 2000. Oxidative stress promotes specific protein damage in *Saccharomyces cerevisiae*, *Journal of Biological Chemistry*, 275: 27393–27398.
- 59) Klaassen, C. D. 1996. Casarett and Doull's Toxicology: The Basic Science of Poisons, *McGraw Hill*, NY, NY 41. 1996.
- 60) Forman, H. J., Liu, R., and Shi, M. M. 1990. Glutathione synthesis in oxidative stress," In: Packer, L., Fuchs, J. (Eds.), *Biothiols in Health and Disease*. Marcel Dekker, NY, NY, pp. 189–211.
- 61) Sies, H. 1999. Glutathione and its role in cellular functions, *Free Radical Biology and Medicine*, 27: 916–921.
- 62) Sunagawa, S., Choi, J., Forman, H. J., and Meidna, M. 2008. Hyperthermic stress-induced increase in the expression of glutamate-cysteine ligase and glutathione levels in the symbiotic sea anemone *Aiptasia pallida*. *Comparative Biochemistry and Physiology Part B*, 151: 133–138.
- 63) Masella, R., and Mazza, G. (Eds.). 2009. *Glutathione and Sulfur Amino Acids in Human Health and Disease*, John Wiley & Sons, Inc., Hoboken, NJ, p. 98.
- 64) Schwarz, J. A., Mitchelmore, C. L., Jones, R., O'Dea, A., and Seymour, S. 2013. Exposure to copper induces oxidative and stress responses and DNA damage in the coral *Montastraea franksi*. *Comparative Biochemistry and Physiology Part C*, 157: 272–279.
- 65) Mitchelmore, C. L., and Hyatt, S. 2004. Assessing DNA damage in cnidarians using the Comet assay, *Marine Environmental Research*, 58: 707–711.
- 66) Sarkar, A., Gaitonde, D. C. S., Sarkar, A., Vashistha, D., D'Silva, C., and Dalal, S. G. 2008. Evaluation of impairment of DNA integrity in marine gastropods (*Cronia contracta*) as a biomarker of genotoxic contaminants in coastal water around Goa, West coast of India, *Ecotoxicology and Environmental Safety*, 71: 473–482.
- 67) Bopp, S. K., Abicht, H. K., and Knaue, K. 2008. Copper-induced oxidative stress in rainbow trout gill cells, *Aquatic Toxicology*, 86: 197–204.
- 68) Almeida, E. A., Bainy, A. C. D., Loureiro, A. P. M., Martinez, G. R., Miyamoto, S., Onuki, J., Barbosa, L. F., Garcia, C. C. M., Prado, F. M., Ronsein, G. E., Sigolo, C. A., Brochini, C. B., Martins, A. M. G., Medeiros, M. H. G., and Mascio, P. 2007. Oxidative stress in *Perna perna* and other bivalves as indicators of environmental stress in the Brazilian marine environment: antioxidants, lipid peroxidation and DNA damage, *Comparative Biochemistry and Physiology Part A*, 146: 588–600.
- 69) Itziou, A., Kaloyianni, M., and Dimitriadis, V. K. 2011. Effects of organic contaminants in reactive oxygen species, protein carbonylation and DNA damage on digestive gland and haemolymph of land snails, *Chemosphere*, 85: 1101–1107.

- 70) Mitchelmore, C. L., and Chipman, J. K. 1998. DNA strand breakage in aquatic organisms and the potential values of the comet assay in environmental monitoring, *Mutation Research*, 399: 135-147.
- 71) Sandrini, J. Z., Bianchini, A., Trindade, G. M., Nery, L. E. M., and Marins, L. F. F. 2009. Reactive oxygen species generation and expression of DNA repair-related genes after copper exposure in zebrafish (*Danio rerio*) ZFL cells, *Aquatic Toxicology*, 95: 285- 291.
- 72) Rainbow, P. S. 2002. Trace metal concentrations in aquatic invertebrates: why and so what? *Environmental Pollution*, 120: 497-507, 2002.
- 73) Cook, C. B., D'Elia, C. F., and Muller-Parker, G., "Host feeding and nutrient sufficiency for zooxanthellae in the sea anemone *Aiptasia pallida*. 1988. *Marine Biology*, 98: 253-262.
- 74) Jones, R. J., Kildea, T., and Hoegh-Guldberg, O. 1999. PAM chlorophyll fluorometry: a new in situ technique for stress assessment in Scleractinian corals, used to examine the effect of cyanide from cyanide fishing, *Marine Pollution Bulletin*, 38: 864-874.
- 75) Nielsen, H. D., Brown, M. T., and Brownlee, C. 2003. Cellular responses of developing *Fucus serratus* embryos exposed to elevated concentrations of Cu<sup>2+</sup>, *Plant Cell Environment*, Vol. 26: 1737-1747.
- 76) Bielmyer, G.K., Grosell, M., Bhagooli, R., Baker, A. C., Langdon, C., Gillette, P., and Capo, T. R. 2010. Differential effects of copper on three species of scleractinian corals and their algal symbionts (*Symbiodinium* spp.), *Aquatic Toxicology*, 97: 125-133.
- 77) Brown, B. E. 2000. The significance of pollution in eliciting the „bleaching" response in symbiotic cnidarians, *International Journal of Environment and Pollution*, 13:392-415.
- 78) Schreiber, U. 2004. Pulse-amplitude-modulation (PAM) fluorometry and saturation pulse method: an overview. In: Papageorgiou, G.C., Govindjee (Eds.), *Chlorophyll a Fluorescence: A Signature of Photosynthesis*. Springer, Amsterdam, pp. 279-319.
- 79) Bielmyer, G. K., Grosell, M., Bhagooli, R., Baker, A. C., Langdon, C., Gillette, P., and Capo, T. R. Assessing effects of copper exposure on three species of scleractinian corals: Implications for coral health, in press.
- 80) Harland, A. D., and Nganro, N.R. 1990. Copper uptake by the sea anemone *Anemonia viridis* and the role of zooxanthellae in metal regulation, *Marine Biology*, 104: 297-301.
- 81) Peters, E. C., Gassmann, N. J., Firman, J. C., Richmond, R. H., and Power, E. A. 1997. Ecotoxicology of tropical marine ecosystems," *Environmental Toxicology and Chemistry* 16: 12-40.
- 82) Jones, R. J. 2004. Testing the „photoinhibition" model of coral bleaching using chemical inhibitors, *Marine Ecology Progress Series*, 284: 133-145.
- 83) Melo, K. V., and Amaral, F. D. 2005. Ampliacao da distribuicao das anemonas-do-mar (Cnidaria, Actiniaria) no estado de Pernambuco. Brasil, *Tropical Oceanography*, 33:19-31.
- 84) Rizzini-Ansari, N. 2009. Evaluation of the sea anemone *Bunodosoma caissarum* as a bioindicator species of THg contamination in Guanabara Bay: a comparison with the mussel *Perna perna*. Senior Thesis, Faculdade de Oceanografia, Universidade do Estado do Rio de Janeiro, 46p (In portuguese).
- 85) Rizzini Ansari, N., et al. 2015. Mercury distribution, methylation and volatilization in microcosms with and without the sea anemone *Bunodosoma caissarum*. *Mar. Pollut. Bull.* <http://dx.doi.org/10.1016/j.marpolbul.2014.12.049>