

# Study of $\alpha$ -1 Antitrypsin Serum and its Effects on Chronic Inflammation in Diabete Patients

Somayyeh Heidary<sup>1</sup>, Ahmad Riahi<sup>2\*</sup>

Department of Cell and Molecular Biology, Universiti Putra Malaysia, Serdang, Malaysia  
Department of Biology, Payam Noor University, Tehran, Iran

Received: 05 January 2018

Accepted: 18 February 2018

Published: 06 March 2018

## Abstract

Pathogenesis of type 2 as well as type 1 diabetes mellitus is observed to be closely associated with acute phase response which is predominately cytokine-mediated. In this present study I try to test this hypothesis by estimating circulating  $\alpha$ 1 antitrypsin (AAT) in freshly diagnosed type 1 (T-1), freshly diagnosed type 2 (T-2) as well as type 2 diabetic patients under oral hypoglycemic drugs for duration of at least five years. AAT is considered as a prominent member of acute phase protein and very important tools for diagnosis for low grade chronic inflammatory reaction. Thirty normal controls to match the age and sex of the test groups were also studied. The level of this parameter was also correlated with their random plasma glucose values and BMI. The value of AAT significantly elevated in the T-2 patients ( $p < 0.0001$ ) in comparison with the controls. In case of T-1 patients the level of AAT found mildly elevated as  $p$  value has marginally significant value of 0.0002. Interestingly in either of the types, no correlation was found with the degree of hyperglycemia or BMI. By the above results and findings it can be definitely postulated that a low grade inflammatory process is surely associated in the pathogenesis of type 2 diabetes. But for type 1 diabetic patient the result is contradictory. This can be further explored for further diagnosis, management and follow up.

**Keywords:**  $\alpha$ 1 antitrypsin; acute phase proteins; acute phase reactants; low grade chronic inflammation; diabetes mellitus;

## How to cite the article:

S. Heidary, A. Riahi, Study of  $\alpha$ -1 Antitrypsin Serum and its Effects on Chronic Inflammation in Diabete Patients, Medbiotech J. 2018; 2(1): 01-07, DOI: 10.22034/mbt.2018.60861

## 1. Introduction

Alpha 1-Antitrypsin (AAT) is an acute phase reactant with antiprotease activity (1) which act as a serine protease inhibitor and recently has been focused for prevention for development of Type 1 diabetes, for prolongation of islet allograft survival and for inhibition of apoptosis of pancreatic B-cell in vivo (2). In addition to its antiprotease activity, AAT has likewise been shown to have other biological effects, including the ability to modulate both inflammation and apoptosis (3,4). AAT was isolated and described in 1955. Interest in this glycoprotein has centered on its genetic polymorphism and the disease associated with its biological function as a protease inhibitor. AAT is a single chain protein of 394 amino acids which contains three oligosaccharide chains. It is the major component (>90%) of the  $\alpha$ -1 fraction of human plasma. It has a molecular weight of  $\sim$ 55000, pI of 4.8 and contains

10-12% carbohydrate. It is synthesized by hepatocytes and macrophages and is the principal serine protease inhibitors of human plasma (5). The name AAT is almost a misnomer since AAT is relatively inactive towards trypsin, although responsible for 90% of serum antitrypsin activity. As a protease inhibitor AAT acts against chymotrypsin, kallikrein, rennin, urokinase, plasmin and possibly thrombin but greater clinical significance regarding inhibitory activity is directed against neutrophil elastase and collagenase by formation of complex with them. The inhibitory activity of AAT is maximal at the neutral to slightly alkaline pH of blood. At pH 4.5 its activity is negligible. When AAT acts as an inhibitor of intestinal enzyme such as trypsin, this pH dependency is less. It is more when it plays in the respiratory tract (6). AAT has significant microheterogeneity (7). Depending on buffer pH and type of support of electrophoresis, a number of

bands may be seen. At least 75 polymorphic forms occur, many of which can be separated by electrophoresis. Even after sialidase treatment, heterogeneity remains unaltered which indicate that AAT phenotypes are probably not due to difference in sialic acid moieties. Typing of multiple forms is empirically based on differences in electrophoretic mobilities (8). Some 33 allotypes have been described. About 95% genotype is PiMM (homozygotic for M protease inhibitor) and is called M protein. Two other proteins Z and S are found in genotypes PiZZ, PiSZ, PiSS, PiMZ and PiMS. Z and S protein differ from one another and from M protein by only one amino acid residue. Circulating level of M protein is directly related to ability to inhibit protease. If normal activity in M phenotype is taken as 100%, the level of activity in ZZ is 15%, in SS 60%, in MZ

57.5% and in MS 80%. A rare null genotype (Pi-) produce no AAT properties at all (9). The function of AAT is to neutralize lysosomal elastase released upon phagocytosis of particles by polymorphonuclear leukocytes. Being a small molecule AAT can pass from capillaries into tissue fluid, bind protease and pass back into intravascular fluid. It may even transfer bound protease to  $\alpha$ -2 macroglobulin, which because of its large size cannot leave the intravascular compartment. By this mechanism protease is rapidly transported to the reticuloendothelial system for rapid degradation.

There is increasing evidence that an ongoing cytokine induced acute phase response which is sometimes called low grade inflammation, but part of a widespread activation of the innate immune system, is closely involved in the pathogenesis of type 2 diabetes mellitus and associated complications such as dyslipidemia and atherosclerosis. Elevated circulating inflammatory markers such as C-reactive protein and interleukin-6 (IL-6) predict the development of type 2 diabetes and several drugs with anti-inflammatory properties such as aspirin and thiazolidinediones lower both acute phase reactants and glycemia and possibly decreased the development of type 2 diabetes. Among the risk factors for type 2 diabetes, which is also known to be associated with active innate immunity, are age, inactivity, certain dietary components, smoking, psychological stress and low birth weight. Other features of type 2 diabetes such as fatigue, sleep disturbance and depression are likely to be at least partly due to hypercytokinemia and activated innate immunity (10).

A major uncertainty is whether hyperglycemia is a main determinant of the inflammation in type 2 diabetes mellitus- there is evidence for and against. Cross sectional studies of type 2 diabetes mellitus show that acute phase reactant like C reactive protein (CRP) and Interleukin-6 (IL-6) are significantly correlated with blood glucose

concentration or glycated hemoglobin percentage (11). Lowering of blood glucose levels in type 2 diabetic patient are accompanied by reduced levels of inflammatory markers (12). Recent finding indicates that acute hyperglycemia in nondiabetic and Impaired glucose tolerance (IGT) subjects, elevate plasma IL-6 and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) concentration higher and longer in an individual with IGT and when glucose was given as pulse (13). By infusion of the antioxidant glutathione the effect was abolished, which suggests that hyperglycemia induced cytokine production is mediated by reactive oxygen species.

Subcutaneous and intraabdominal adipose tissue is a major source of TNF- $\alpha$  and IL-6 production (14). This raise the question of whether the acute phase reaction of type 2 diabetes is mainly secondary to obesity. In a recent study in which a case and control subjects were matched by body mass index (BMI) and waist circumference, neither CRP nor IL-6, predicted the development of type 2 diabetes, although lowered level of adiponectin did. Thus a hypothesis was suggested that because inflammatory markers are associated with obesity, they only indirectly predict diabetes and act as surrogate markers of hypo adiponectinemia (15).

Atherosclerosis is another co-segregate of type 2 diabetes which is strongly associated with acute phase response in its own right (16). Present evidence support the notions that atherosclerosis develops in parallel with type 2 diabetes (17) with both conditions showing the common antecedent of activated innate immunity, but like hyperglycemia and possibly some other manifestation of type 2 diabetes such as obesity, microangiopathy once present, would presumably further enhance inflammation.

Where there are studies on type 2 diabetes mellitus agreeing on the fact that acute phase proteins especially AAT are increased, studies in type 1 patients remains contradictory. It was found that acute phase markers are not elevated or may be mildly elevated in type 1 subjects who had the same degree and duration of hyperglycemia as type 2 subjects. At the same time, type 1 patients are at the same risk of developing atherosclerosis as the type 2 patient. Hence it was speculated that specifically diabetes related factors (possibly glucose) would need to be additionally sensitize the arteries to cytokines and other atherogenic factors such as hypercholesterolemia. It has also been shown that Type 1 diabetic patients have marginally higher plasma concentrations of AAT, suggesting the potential role of AAT in the pathogenesis of Type 1 diabetes. We have investigated the role of AAT as an inflammatory marker in pathogenesis of type 2 diabetes mellitus (18).

## 2. Materials and Method

### 2.1 Aims and objective

1. To detect the elevation of AAT, if any, in newly diagnosed untreated type 1 diabetes mellitus patients, in newly diagnosed untreated type 2 diabetes mellitus patients and in patient of type 2 diabetes mellitus under treatment for at least five years. 2. Compare the levels in newly diagnosed untreated type 2 diabetes mellitus patient with type 2 diabetes mellitus patients under treatment for at least 5 years. 3. Compare the level of this inflammatory marker of newly diagnosed untreated type 1 diabetes mellitus with newly diagnosed untreated type 2 diabetes mellitus patients.

### 2.2 Participants

Subjects were selected from various clinics and hospitals in Mangalore, India. Height and weight of all subjects were recorded and body mass index was calculated. None of the ninety two volunteers were alcoholics or smokers. The participants did not suffer from chronic inflammatory diseases like asthma, chronic bronchitis, and rheumatoid arthritis as was ascertained by clinical history. The study was approved by institutional ethical committee of Kasturba Medical College, Mangalore, India.

### 2.3 Materials

5 ml blood was collected in plain bottle. Informed consent was taken from the individual subjects prior to blood collection. Blood was taken from antecubital vein of the subjects and AAT assay in serum was carried out by the method of Sundaresh et al (19).

### 2.4 Principle of the test

The Proteolytic enzyme trypsin hydrolyses casein, with the formation of smaller peptides. The enzyme reaction after suitable interval of time is arrested by the addition of TCA which precipitated the protein, but the peptides are soluble in the acid. The TCA soluble fragments are a measure of Proteolytic activity of the enzyme. When the inhibitor is added to the preincubated mixture, it prevents the release of peptides by the Proteolytic enzymes. Thus the estimation of TCA soluble components in the presence and absence of inhibitor is a measure of inhibitory activity against Proteolytic enzymes. The TCA soluble fragments are analyzed by the method of Lowry et al. The final color formed is a result of Biuret reaction of the peptides with copper ions in alkali and reduction of the phosphomolybdic acid reagent by the presence of tyrosine and Tryptophan which are present in the treated peptides.

### 2.5 Procedure

The mixture was preincubated at 37°C for 10 minutes. The enzyme reaction was started by the addition of 1 ml of 2% casein. After 20 minutes incubation at 37°C, the enzyme reaction was arrested by the addition of 3ml of 5% TCA. After standing for 30 minutes it was centrifuged for 15 minutes.

Contents	Enzyme blank	Enzyme control	Test blank	Test
Buffer	0.5 ml	0.5ml	0.5 ml	0.5 ml
Water	0.5ml	0.3ml	0.4 ml	0.2 ml
Diluted sample	-	-	0.1 ml	0.1 ml
Diluted enzyme 1: 300	-	0.2ml	-	0.2 ml

To 1 ml of supernatant, 1 ml of water and 4 ml of alkaline copper reagent were added. After 10 minutes, 0.4 ml of Folin reagents was added. Blue color developed was measured at 540 nm after 13 minutes.

The inhibitor concentration was calibrated in term of mgs of Trypsin inhibited, from the difference in the enzyme activity with and without sample. One unit of inhibitor activity is defined as 1 mg of enzyme inhibited under the assay condition.

### 2.6 Calculation

The data was analyzed by the students't test and the ANOVA test. Pearson's coefficient was applied for co relational analysis.

## 3. Results

The aim of the study was to examine the level of AAT as an inflammatory marker in pathogenesis in diabetes mellitus. The mean age (range), body mass index (BMI) and values of random blood sugar (RBS) are presented in table 1. The control group participants were so chosen as to cover the age range of the test groups. Table 2 lists the values of AAT in all four groups as mean  $\pm$  SD. Table 3 denoted the comparison between different groups and significance levels (p values). In newly diagnosed type 2 diabetic patients (Group II) show higher level of AAT in compare to control group(Group IV) as depicts the significant p value(< 0.0001\*). It's also detect that the level of the AAT is moderately higher in newly diagnosed type I(Group II) in compare to control (Group IV) as denoted by marginally significant p value (0.0002). It is also shown that level of AAT is even significantly higher (Group III) even after treatment by oral hypoglycemic drugs in compare to control group. (Group IV)

**Table 1.** The anthropometric data of the subjects participated in the study are presented in

	Group I(n=12) (Mean $\pm$ SD)	Group II (n=25) (Mean $\pm$ SD)	Group III (n=25) (Mean $\pm$ SD)	Group IV(n=30) (Mean $\pm$ SD)
Age (yrs)	18.33 $\pm$ 7.64	48.22 $\pm$ 7.11	51.32 $\pm$ 7.56	44.97 $\pm$ 15.06
BMI	19.50 $\pm$ 1.23	24.03 $\pm$ 1.46	24.20 $\pm$ 2.40	21.75 $\pm$ 2.27
RBS	338.25 $\pm$ 50.97	193.26 $\pm$ 35.30	93.61 $\pm$ 33.65	94.20 $\pm$ 7.00

Group I = Type 1 diabetes mellitus patient (newly diagnosed)

Group II = Type 2 diabetes mellitus patient (newly diagnosed)

Group III = Type 2 diabetes mellitus patient (under treatment)

for at least 5 years) Group IV = Control  
 n = number of subjects SD = Standard Deviation  
 BMI= Body Mass Index  
 RBS= Random Blood Sugar

**Table 2.** The compare of mean value of AAT.

Serum Level (mg/dl)	Group I (Mean ± SD)	Group II (Mean ± SD)	Group III (Mean ± SD)	Group IV (Mean ± SD)
AAT (mg/dl)	495.70 ± 32.77	562.16 ± 63.00	519.38 ± 47.80	350.48 ± 114.07

Group I Group II Group III Group IV

SD = Standard Deviation

Group I = Type 1 diabetes mellitus patient (newly diagnosed)

Group II = Type 2 diabetes mellitus patient (newly diagnosed)

Group III = Type 2 diabetes mellitus patient (under treatment for at least 5 years)

Group IV = Control

**Table 3.** Comparison of level of AAT (mg/dl) between different groups in Table-3 (p value < 0.05 is considered significant).

Comparison between groups	Level (mg/dl)	Level (mg/dl)	p value
Comparison between Group I and Group IV	495.70 ± 32.77 (I)	350.48 ± 114.07 (IV)	0.0002*
Comparison between Group II and Group IV	562.16 ± 63.00 (II)	350.48 ± 114.07 (IV)	< 0.0001*
Comparison between Group III and Group IV	519.38 ± 47.80 (III)	350.48 ± 114.07 (IV)	< 0.0001*
Comparison between Group I and Group II	495.70 ± 32.77 (I)	562.16 ± 63.00 (II)	0.003
Comparison between Group II and Group III	562.16 ± 63.00 (II)	519.38 ± 47.80 (III)	0.03

#### 4. Discussion

The aim and objective of this study was to determine whether low grade chronic inflammation as a pathogenic cause in type 1 and type 2 diabetes mellitus or not. AAT levels were determined as a prominent inflammatory marker. In the twenty-five newly diagnosed type 2 patients the level of  $\alpha$ 1antitrypsin was found to be significantly increased as compared to control (Table 3). The role of chronic low grade inflammation in the pathogenesis of type 2 diabetes seems possible beyond doubt. At the same time its role in type 1 diabetes cannot be completely ruled out.

The underlying mechanism for the augmented acute phase response is not well understood and the stimulus for the response is unknown. Characteristic changes in liver protein synthesis occur in response to tissue damage from infection or injury known collectively as the acute phase response (20). In this setting, increased hepatic production of a group of so-called "acute phase reactant" proteins. AAT, together with creatinine protein, ferritin, complement factors, and others are significantly up regulated by the result of IL-6-, TNF $\alpha$ -, and IL-1-driven signal transducer and activator of transcription 3 (STAT3) and nuclear factor kappa B (NF- $\kappa$ B) p65 signaling, while cortisol-binding globulin, transferrin, and albumin are decreased which are considered as negative acute phase proteins(21,22).

Circulating blood AAT levels in the acute phase have been found to increase three- to four fold (23). In addition to the acute phase response, AAT levels

can also increased by other inflammatory settings including pregnancy, trauma, surgery, malignancy, and treatment with oral contraceptives(24,28). Finally, there is some evidence that circulating granulocytes could contribute to local AAT levels through the transcription, storage, and release of AAT following migration into tissues(29-31). While some studies have demonstrated that AAT is stored in secretory vesicles within neutrophils (30,31), more recent data suggests that approximately 80 % of neutrophil-associated AAT is localized to the cell membrane within lipid rafts, where it co-localizes with Fc fragment IgG receptor IIIb (Fc $\gamma$ RIIIb) (29).

Intriguingly, many of the anti-inflammatory characters of AAT appear to be not dependent of its antiprotease activity. Churg and colleagues shown that oxidized AAT, lacking in antiproteolytic capacity, attenuated silica-induced increases in lung monocyte chemoattractant protein 1(MCP-1) expression, macrophage inflammatory protein- 2  $\alpha$  (MIP-2  $\alpha$ ), activation of NF- $\kappa$ B, and associated neutrophilic migration into the lung (32-33). More recently, Jonigk and colleagues demonstrated that a recombinant form of AAT lacking elastase inhibitory function exhibited anti-inflammatory properties in lipopolysaccharide (LPS) challenged mice, decreased infiltration of neutrophils, decreased TNF- $\alpha$ , decreased lung tissue expression of DNA damage inducible transcript 3 protein (DDIT3) and X-box binding protein-1 (XBP-1) as well as in freshly isolated human blood neutrophil (decreased TNF $\alpha$  and IL-8 secretion ex vivo)(32-33). While antiprotease activity may not be necessary for AAT-mediated effects on inflammation, proper protein glycosylation does seem to be significant. Hyperglycemia and insulin resistance could promote inflammation and inflammation may be a factor linking diabetes mellitus to the development of atherosclerosis. Elevated glucose levels promote inflammation by increasing oxidative stress by increased formation of tumor necrosis factor (TNF) kappa B (34). In this study, the mean BMI was found to be 19.5 ± 1.23 in type 1 patient and 24.03 ± 1.46 in type 2 patients. No correlation was found between BMI and acute phase reactants. Hence it can be summarized that there could be multiple pathways involved in the activation of the innate immunity system and much work needed to be done to establish either a casual role in the development of mainly type 2 diabetes and could be type 1 diabetes also.

Having demonstrated that there is an inflammatory process going on in type 2 diabetes, we next thought of estimating inflammatory markers in patients on treatment ( for at least 5 years) with oral hypoglycemic drugs. Many of the drugs have been shown to have anti-inflammatory effects. Statin drugs inhibits HMG-CoA reductase and prevent

atherosclerosis and inhibit the acute phase response by diminishing the deposition of low density lipoprotein (LDL) particles rich in cholesterol and phospholipids in macrophages and smooth muscle cells (35). Statins were found to reduce CRP levels and did not correlate with the reduction of the lipid levels suggesting that in addition to their ability to reduce LDL, statins may also inhibit the acute phase response (36). Freeman DJ et al showed that statin therapy also prevent diabetes mellitus. Pravastatin therapy in the West of Scotland Coronary Prevention Study resulted in a 30% reduction of risk of developing type 2 diabetes(37). Salicylates in high doses have been known to lower glycosuria in diabetic patients (38). Although earlier studies were contradictory, these studies has used lower aspirin doses (<3gm/day) and therapeutic duration was only for a few days. Hundal RS (39) reported that high doses of aspirin (7 gm/day) for 2 wks caused 25% reduction in fasting plasma glucose, 50% reduction in triglyceride and 15 % reduction of CRP concentration independently of the changes in the plasma insulin concentration. Another widely used oral hypoglycemic agents thiazolidinedions (Glitazone) are peroxisome proliferators activated receptor  $\gamma$  ( PPAR  $\gamma$ ) agonist that have been regarded as insulin sensitizers through mechanisms such as altered transcription of insulin sensitive genes controlling lipogenesis, adipocytes differentiation, fatty acid uptake and GLUT 4 ( Glucose Transporter 4) expression. They also have an anti-inflammatory action inhibiting cytokine production, macrophage activation and reducing CRP as well as WBC count in type 2 diabetic subjects (40-43).

Angiotensin Converting Enzyme Inhibitors (ACE inhibitors) are also known to decrease insulin resistance in either type 1 or type 2 diabetic patients with concomitant hypertension (44) Torloni E et al demonstrated improved glycemic control in patients with arterial hypertension and type 2 diabetes using ACE inhibitors (45) Insulin has a potent anti-inflammatory activity. Insulin was found to be a rapid nonspecific and dose dependent inhibitors of the cytokine and glucocorticoids stimulation of acute phase protein, gene expression and exerted effect at the transcriptional levels. Insulin inhibition applied to all cell cytokines tested but to various degrees depending upon the particular acute phase gene (46).

In this study, of the 25 type 2 diabetic patients on treatment for at least 5 yrs, 8 patient were in sulfonylurea-metformin combination, 7 were on glitazone, 6 were on sulfonylurea alone, 2 were on glitazone-metformin combination and 2 were on metformin alone. In newly diagnosed untreated type 2 diabetic group (Group II) the levels of AAT is statistically elevated in compare to the control group (Group IV) which used to demonstrate that

type 2 diabetes trigger the process of inflammation. When the level of AAT in treated group (Group III) compared with untreated group (Group II) not shown any statistically significance p value (0.03), that may be demonstrated that oral hypoglycemic therapy don't have any significant effect to reduce the inflammatory condition triggered by type 2 diabetic patients. Irrespective of oral hypoglycemic drug used for the treatment, treated group (Group III) showed significantly higher AAT levels comparable to the control group (Group IV).

## 5. Conclusion

What causes the innate immunity activation? Is it a cause or an effect of diabetes? What is the role of hyperglycemia? Is it associated with other complications of diabetes and if so, how? - are a few of the question which need to be addressed by intensive research. The mechanisms could be multifactorial and complex. A few hypothesis have been postulated which are still wanting.

## References

1. Laurell CB, Jeppson JO: Protease inhibitors in plasma. In; The plasma Proteins, F Putamen, Ed New York; Academic press 1975; Vol 1, 2nd Edi:229
2. Sandström CS, Ohlsson B, Melander O, Westin U, Mahadeva R, Janciauskiene S. An association between type 2 diabetes and alpha-antitrypsin deficiency, *Diabet Med.* 2008; Nov; 25(11):1370-3.
3. Petrache I, Fijalkowska I, Medler TR, Skirball J, Cruz P, Zhen L, Petrache HI, Flotte TR, Tudor RM.  $\alpha$ -1 antitrypsin inhibits caspase-3 activity, preventing lung endothelial cell apoptosis. *Am J Pathol.* 2006; 169:1155-66.
4. Janciauskiene SM, Bals R, Koczulla R, Vogelmeier C, Köhnlein T, Welte T. The discovery of  $\alpha$ 1-antitrypsin and its role in health and disease. *Respir Med.* 2011; 105:1129-39.
5. Murry RK, Granner DK, Mayer PA, Rodwell VW. Plasma protein, immunoglobulin and blood coagulation. In; Harpers Biochemistry. McGraw Hill.2002; 25th edition :744
6. Tietz NW: Amino acids and proteins: In; Textbook of clinical chemistry: WB Saunders Company, 1986; 519618.
7. Johnson AM: Genetically determined variations in plasma protein, In; Protein abnormalities: Proteins in body fluid, Amino acids and tumor markers: Diagnostic and clinical aspects., Se Ritzmann and LM Killingsworth, Ed s New York: Alan RLiss, Inc.1983; vol1: 53-100
8. Jeppsson JO, Franzen B. Typing of genetic variants of  $\alpha$ 1 antitrypsin by electrofocussing. *Clin Chem* 1982; 28:219-25
9. Talamo RC, Largley CE and Reed CE. $\alpha$ -1 antitrypsin deficiency: A variant with no detectable  $\alpha$ -1 antitrypsin. *Science* 1973; 181:70

10. Pickup JC: Inflammation and activated innate immunity in the pathogenesis of type 2 diabetes. *Diabetes care*. 2004; 27 (3) : 813
11. Rodriguez-Moran M, Guerrero-Romero F. Increased level of C-reactive protein in noncontrolled type 2 diabetic subjects. *J Diabetes Complications*. 1999; 13: 211-5
12. Ceriello A, Mercuri F, Fabbro D, Giacomello R, Stel G, Taboga C, Tonutti L, Motz E, Damante G. Effects of intensive glycemic control on fibrinogen plasma concentration in patient with type 2 diabetes: relation with  $\beta$ -fibrinogen genotype. *Diabetologia*. 1998; 41: 1270-3.
13. Esposito K, Nappo F, Marietta R, Giugliano G, Giugliano L, Ceriello A, Giugliano D. Inflammatory cytokines concentration are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation*. 2002; 106: 2067-72.
15. Mohamed Ali V, Goodrick S, Rawesh A, Mile JM, Katz DR, Yudkin JS, Coppack SW. Human subcutaneous adipose tissue releases IL-6 but not TNF- $\alpha$  in vivo. *J Clin Endocrinol Metab*. 1997; 82: 4196-200
16. Krakott J, Funahashi T, Slehouwer CD, Schalkwijk CG, Tanaka S, Matsuzawa Y, Kobes S, Tataranni PA, Hanson RL, Knowler WC, Lindsay RS. Inflammatory markers, adiponectin and risk of type 2 diabetes in the Pima Indian. *Diabetes Care*. 2003; 26:1745-51.
17. Smart J, George AG, Davies AJ, Auklund A, Hurlow RA: Haematological stress syndrome in atherosclerosis. *J Clin Pathol*. 1981; 34:464-7.
18. Pickup JC, Mattlock MB. Activation of innate immune system as a predictor of cardiovascular mortality in type 2 diabetes mellitus. *Diabet Med* 2003; 20: 723-6.
19. Crook MA, Tutt P, Simpson H, Pickup JC: Serum sialic acid and acute phase protein in type 1 and type 2 diabetes. *Clin Chim Acta*. 1993; 219 : 131-8.
20. Sundaresh CS, Aroor AR and Pattabiraman TN. Comparative study of amidolytic and caseinolytic methods for the determination of urinary trypsin inhibitor. *Indian J Med Res*. 1978; 68: 341-4.
21. Baumann H, Gauldie J. The acute phase response. *Immunol Today*. 1994;15:74-80.
22. Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *NEJM*. 1999;340:448-54.
24. Quinton LJ, Blahna MT, Jones MR, Allen E, Ferrari JD, Hilliard KL, Zhang X, Sabharwal V, Algül H, Akira S, Schmid RM, Pelton SI, Spira A, Mizgerd JP.
25. Hepatocyte-specific mutation of both NF- $\kappa$ B RelA and STAT3 abrogates the acute phase response in mice. *J Clin Invest*. 2012;122:1758-63.
26. Perlmutter DH, May LT, Sehgal PB. Interferon beta 2/interleukin 6 modulates synthesis of alpha 1antitrypsin in human mononuclear phagocytes and in human hepatoma cells. *J Clin Invest*. 1989; 84:138-44.
27. Mendenhall HM. Effect of oral contraceptive on serum protein concentrations. *Am J Obstet Gynecol*. 1970;106:750-3.
28. Lisowska-Myjak B, Pachecka J. Antigenic and functional levels of alpha-1-antitrypsin in serum during normal and diabetic pregnancy. *Eur J Obstet Gynecol Reprod Biol*. 2003; 106: 31-5.
29. Crystal RG. Alpha 1-antitrypsin deficiency, emphysema, and liver disease. Genetic basis and strategies for therapy. *J Clin Invest*. 1990; 85:1343-52.
30. Kishore N, Rizvi V, Kishore N, Sharma BB. Effects of oral contraceptive on alpha I-antitrypsin activity. *J Obstet Gynaecol India*. 1979; 29:259-60.
31. Palmer PE, Christopherson WM. Alpha1-antitrypsin, protein marker in oral contraceptive associated hepatic tumors. *Am J Clin Pathol*. 1977; 68:736-9.
32. Bergin DA, Reeves EP, Meleady P, Henry M, McElvaney OJ, Carroll TP, Condron C, Chotirmall SH, Clynes M, O'Neill SJ, McElvaney NG.  $\alpha$ -1 Antitrypsin regulates human neutrophil chemotaxis induced by soluble immune complexes and IL-8. *J Clin Invest*. 2010; 120:4236-50.
33. Johansson B, Malm J, Persson T, Janciauskiene S, Andersson P, Carlson J, Egesten A. Alpha-1-antitrypsin is present in the specific granules of human eosinophilic granulocytes. *Clin Exp Allergy*. 2001; 31:379-86.
34. Pääkkö P, Kirby M, du Bois RM, Gillissen A, Ferrans VJ, Crystal RG. Activated neutrophils secrete stored alpha 1-antitrypsin. *Am J Respir Crit Care Med*. 1996; 154:1829-33.
35. Jonigk D, Al-Omari M, Maegel L, Müller M, Izykowski N, Hong J, Hong K, Kim S-H, Dorsch M, Mahadeva R, Laenger F, Kreipe H, Braun A, Shahaf G, Lewis EC, Welte T, Dinarello CA, Janciauskiene S. Anti-inflammatory and immunomodulatory properties of  $\alpha$ 1antitrypsin without inhibition of elastase. *Proc Natl Acad Sci U S A*. 2013; 110: 15007-12.
36. Churg A, Dai J, Zay K, Karsan A, Hendricks R, Yee C, Martin R, MacKenzie R, Xie C, Zhang L, Shapiro S, Wright JL. Alpha-1-antitrypsin and a broad spectrum metalloprotease inhibitor, RS113456, have similar acute anti-inflammatory effects. *Lab Invest*. 2001; 81:1119- 31.
37. Brownlee M. Biochemistry and molecular cell biology of diabetic complication. *Nature* .2001; 414: 813-20
38. Manford RS. Statins and the acute phase response. *N Engl J Med*. 2001; 344: 2016-8.
39. Sparrow CP, Burton CA, Hernandez M. Simvastatin has anti-inflammatory and antiatherosclerosis activities in depend of plasma cholesterol lowering. *Arterioscler Thromb Vasc Biol*. 2001; 21: 115-21.

40. Freeman DJ, Norrie J, Satter N. Pravastatin and the development of diabetes mellitus: evidence for a protective treatment in the West of Scotland Coronary Prevention Study. *Circulation*. 2001; 103: 357-62.
41. Yuan M, Konstanopoulos N, Lee J, Hansen L, Li ZW, Karin M, Shoelson SE. Reversal of obesity and diet induced insulin resistance with salicylate or targeted disruption of ikk(beta). *Science*. 2001; 293: 1673-7
42. Hundal RS, Petersen KF, Mayerson AB, Randhawa PS, Inzucchi S, Shoelsen SE, Shulman GI. Mechanism by which high dose aspirin improves glucose metabolism in type 2 diabetes. *J Clin Invest*. 2002; 109: 1321-6.
43. Ricole M, Li AC, Willison TM, Kelly CJ, Glass CK: The peroxizome proliferators activated receptor(gamma) is a negative regulator of macrophage activation: *Nature*. 1998; 391:79-82
44. Haffner SM, Greenberg AS, Westor WM, Chen H, Williams K, Freed MI. Effect of rosiglitazone treatment on nontraditional markers of cardiovascular disease in patients with type 2 diabetes mellitus. *Gradation*. 2002; 106: 679-84.
45. Chu NV, Kong APS, Kim DD, Armstrong D, Baxi S, Deustch R, Caullied M, Mudaliir SR, Reitz R, Hencry RR, Reaven PD. Differential effects of metformin and troglitazone on cardiovascular risk factors with type 2 diabetes. *Diabetes Care*. 2002; 25:542-9.
46. Ebeling P, Teppo AM, Koiestinen HA, Viikari J, Ronnemma T, Nissen M, Bergkulla, Salmela P, Saltevo J, Koivisto VA. Troglitazone reduces hyperglycaemia and selective acute phase proteins in patients with type 2 diabetes. *Diabetologia*. 1999; 42: 1433-8.
47. Pollare T, Lithell H, Berne CA. A comparison of the effects of hydrochlorothiazide and captopril on glucose and lipid metabolism in patients with hypertension. *N Engl J Med*. 1989; 321: 866-72.
48. Torlone E, Britta M, Rambotti AM, Periello G, Santeusanio F, Brunetti P, Bolli GB. Improved glycemic control after long term angiotensin converting enzyme inhibition in subjects with arterial hypertension and type 2 diabetes. *Diabetes Care*. 1993; 16: 1347-52.
49. Campus SP, Baumann H. Insulin is a prominent modulator of the cytokine stimulated expression of acute phase plasma protein gene. *Mol Cell Biol*. 1992; 12(4): 1789-97.